



Research Article

EFFICACY OF DIFFERENT INSECTICIDES AGAINST PINK BOLLWORM *Pectinophora gossypiella* (SAUND.) (LEPIDOPTERA:GELECHIIDAE) ON COTTON CROP IN ECOLOGICAL ZONE OF RAHIM YAR KHAN.

Ashiq Hussin Sanghi¹, Tanweer Ahmed², Muhammad Aslam³, Laila Khalid⁴ and Asma Aslam⁵

¹Senior Subject Matter Specialist (Plant Protection) Adaptive Research Farm, Rahim Yar Khan

²Director (Farms and Trainings) Adaptive Research Rahim Yar Khan

³Senior Subject Matter Specialist (Agronomy) Adaptive Research Farm, Rahim Yar Khan

⁴Assistant Research Officer (PP) Adaptive Research Rahim Yar Khan

⁵ Student of PMAS Arid Agriculture University, Rawalpindi

Abstract

A field experiment was conducted during summer seasons 2016-17 to evaluate the efficacy of different insecticides against pink bollworm *Pectinophora gossypiella* (Saund.) (Lepidoptera:Gelechiidae). Pink bollworm is the major pest of cotton. The larvae feed on the seeds and destroy the fibers of cotton, reducing quality and crop yield.

Six different insecticides Triazophos 40EC @2500ml/ha, Deltamethrin 2.5EC @1000ml/ha, Triazophos + Deltamethrin 40EC+2.5EC @ 2500+1000ml/ha, Gamma cyhalothrin 60CS @250ml/ha, Spinetoram 120SC @300ml/ha and Chlorantraniliprole 20SC @312ml/ha were evaluated with a check plot. Experiment was laid out in RCBD with three replications. Before application of insecticides, the pre treatment observations were taken out, while post treatment observations were taken after 3rd and 7th day of application of insecticides. However, Triazophos + Deltamethrin 40EC+2.5EC @ 2500+1000ml/ha, (89.8%) showed its effectiveness up to 7th day of the spray during both years. The mortality percentage of pink bollworm on cotton was observed in case of Gamma cyhalothrin 60CS as (79.1%), Spinetoram 120SC (73.4%), Deltamethrin 2.5 EC as (65.1%) and Triazophos 40 EC as (59.8%). The effectiveness of insecticides was observed up to 7th day after application during both years 2016-17. Chlorantraniliprole 20SC showed less efficacy (51.6%) during both years.

Keywords: *Pectinophora gossypiella*, pink bollworm, insecticides, mortality percentage.

Introduction

Cotton is the most important crop of Pakistan, cultivated on 2917 thousands hectares and is the source of large amount of foreign exchange, contributing about 5.2% of value added in agriculture and about 1.0 percent of GDP and

contributes about 66% share in national oil production (Anonymous, 2015). In textile manufacturing, it produces seeds with a potential multi product base such as hulls, oil, lint and food for animals (Ozyigit et al. 2007). The insect pest infestation in cotton caused deterioration in lint quality and 10–40% losses in crop yield

(Gahukar, 2006). Pink bollworm, *Pectinophora gossypiella* (Saund.) and the spiny bollworm, *Earias insulana* (Boisd.) are the most serious cotton pests of cotton (Hussein *et al.* 2002). The spiny bollworm larvae damage buds early in the growing season and squares, and bolls later in the season. The reduction in cotton yield was mostly related to the late season infestation with two species and the economic yields are almost impossible to achieve without their chemical control (Younis *et al.* 2007). The pink bollworm (PBW) was described by W.W. Saunders in 1842 from specimens damaging cotton in India. More recent studies suggest the origin of the PBW as the eastern Indian Ocean area bordered on the east by northwestern Australia and on the west by various islands of Indonesia-Malaysia (Common, 1958; Wilson, 1972). Female pink bollworm moths lay eggs singly or, more commonly, in small groups. The eggs of this bollworm are minute and difficult to see without some magnification. Eggs are white when first laid but then turn orange, and later the larval head capsule is visible prior to hatching. Eggs hatch in about three to four days after they are laid (Vennila *et al.*, 2007). Pink bollworm larvae feed on flower buds, flowers, bolls and the seeds within Damage to developing seeds, and the termination of growth results in boll rotting, premature or partial boll opening, reduction of staple length, strength, and increases trash content in the lint (Schwartz, 1983), although 100% crop loss can occur with heavy infestations. Pink bollworms spend the winter as diapausing larvae, then pupate and emerge as adults in spring and early summer (Bariola & Henneberry, 1980). Cotton growers have experienced severe economic losses from the PBW due to reduced yields, low lint quality and the increased costs of insecticides (Watson and Fullerton, 1969; Burrows *et al.*, 1982). When susceptible bolls are available, eggs are laid under the calyx or on bracts and are relatively inaccessible to insecticide deposits. Larvae enter the boll soon after hatching from the eggs and are not killed by insecticides when they are within the boll. Sampling for bolls infested with eggs instead of larvae can reduce insecticide use by $28\pm 35\%$ without any significant loss of yield or lint quality

(Hutchison *et al.*, 1988, 1991). PBW resistance to chlorinated hydrocarbons was reported in Mexico and Texas in the 1950s and 1960s (Lowry and Berger, 1965) and tolerance to certain synthetic pyrethroid compounds has occurred more recently (Haynes *et al.*, 1986, 1987). Insecticide applications are generally terminated in mid- to late September because of the high treatment costs and reduced benefits in the potential yield. High larval populations that occur in late season bolls represent the overwintering diapause generation. Typically, 90% of the total upland cotton bolls produced are set by 15 September. Bolls set after late August to mid-September may not mature or may produce fiber of low quality (Bennett *et al.*, 1967). Also, biotic compound played an important role in pest control, among these compounds spinosad, it is a soil dwelling bacterium called *Saccharopolyspora spinosa*. Spinosad possesses less risk than most insecticides to mammals, birds, fish and beneficial insects. It was used for control of Lepidoptera insect (Temarak, 2007, Ghure *et al.*, 2008 and Gosalwad *et al.*, 2009). For assessing the infestation by the pink and spiny bollworms, 100 green bolls were collected weekly at random from both diagonals of the inner square area for each plot (25 bolls for each replicate) according to the method of Shaaban and Radwan (1974). Integrated pest management (IPM) has been defined as a sustainable approach to managing pests by combining biological, cultural, physical and chemical tools in a way that minimizes economic, health and environmental risks (Greenberg *et al.*, 2012). During the last two decades gossyplure, the sex pheromone of the pink bollworm, is widely used for monitoring and timing of the insecticidal applications. However, due to several reasons, trap catches do not always correspond to actual changes in infestation levels (Henneberry and Clayton 1982, Flint *et al.* 1993). Abd El-Mageed *et al.* (2007) reported that the sequence of betacyfluthrin, malathion and spinosad and the sequence of lufenuron, malathion and spinosad induced the highest reduction (81.04 and 86.08%) in infestation of pink bollworm larvae, respectively. El-Metwally *et al.* (2003) showed that synthetic pyrethroids (Fenprothrin,

esfenvalerate and lambda-cyhalothrin) gave the highest reduction of bollworm infestation followed by IGRs, flufenoxuron, hexaflumzuron mixed with O.P chlorpyrifos, while IGRs alone gave least reduction. In response to this problem effort to control or suppress bollworm damage to growing cotton frequently involve using insecticides of different groups in rotation program which may be useful to delay the resistance problem. Khidr et. al. (1996), El-Sorady et. al. (1998), and Abd El Mageed et. al. (2007) indicated that application of insecticides in sequential use induced higher reduction in larval number as compared with the lower reduction resulting from several applications with the same insecticide alone. So use of chemicals is an essential part of integrated pest management in crop protection measures (Mohyuddin et al. 1997). Non selective use of pesticides leads to water pollution, soil degradation, pest resistance and resurgence and ozone depletion (Naeem et al. 2012).

The pink bollworm is now recorded in nearly all the cotton growing countries of the world, and is a key pest in many of these areas. Keeping in view the economic importance and pest status of pink bollworm, this study was conducted to evaluate and screen out the most effective selective insecticide for the management of pink bollworm on cotton crop.

Materials and Methods

The experiment was conducted at Adaptive Research Farm Rahim Yar Khan on pink bollworm *Pectinophora gossypiella* (Saund.) (Lepidoptera:Gelechiidae) on cotton crop during 2016 and 2017 to test the efficacy of six insecticides viz. Triazophos 40EC @2500ml/ha, Deltamethrin 2.5EC @1000ml/ha, Triazophos + Deltamethrin 40EC+2.5EC @ 2500+1000ml/ha, Gamma Cyhalothrin 60CS @250ml/ha, Spinetoram 120SC @300ml/ha and Chlorantraniliprole 20SC @312ml/ha on cotton variety, MNH-886. The experiment was laid out in Randomized Complete Block Design (RCBD). The insecticides used in the present experiment

were obtained from the local market and were sprayed at field recommended doses when the population of pest reached the Economic Threshold Level (ETL). The ETL for the pink bollworm is considered as 5 live larvae/100 bolls. There were 7 treatments including control, having 3 repeats. The plot size was kept as 20 ft x 60 ft. The plant inspection method was used for sampling the pest population. The field recommended doses of the insecticides as presented in Table 1 were sprayed with hand operated knapsack sprayer having 20 liters capacity fitted with hollow cone nozzle. The control plot remained un sprayed. The sprayer was calibrated using simple water by calculating the amount of water required for spraying on a unit area prior to experiment. All agronomic practices like irrigation, fertilizer applications etc. were kept uniform throughout the experiment on all plots. Pest data was recorded from 10 randomly selected plants. To study the efficacy of different insecticides as mentioned in (Table 1), population of pink bollworm was recorded by the same method 3rd and 7th days after treatment. Crop was kept free from weeds. Mortality of pest was calculated with following Henderson Tilton's formula (1955):

$$\left(1 - \frac{n \text{ in Co before treatment} * n \text{ in T after treatment}}{n \text{ in Co after treatment} * n \text{ in T before treatment}}\right) \times 100$$

Where : n = Insect population , T = treated , Co = control

Data were analyzed statistically with M-stat package and means were compared by DMR test at 5 percent probability level (Duncan, 1955).

Table 1. Different insecticides used against pink bollworm, *Pectinophora gossypiella* (Saund.) with respective doses per hectare.

S. #	Insecticides with formulation	Dose (ml or gm/ha)
1	Triazophos 40EC	2500
2	Deltamethrin 2.5EC	1000
3	Triazophos + Deltamethrin 40EC+2.5EC	2500+1000
4	Gamma Cyhalothrin 60CS	250
5	Spinetoram 120SC	300
6	Chlorantraniliprole 20SC	312

Results and Discussion

Insecticides (Table 1) were sprayed in recommended doses when the population of cotton pink bollworm reached economic threshold level (ETL). Insecticides were dissolved in water to prepare insecticide solutions on vol. / vol. and Wt. / Vol. basis. The crop was sprayed in the morning before 10 a. m. The population of insect pests was recorded 3rd and 7th days after application of insecticides. The effectiveness of various insecticides was considered to be an indirect reflection of pest population in various treatments i.e. lower population of insect pests would represent higher toxicity and vice versa.

The population of cotton pink bollworm was significantly lower ($P < 0.05$) in insecticides treated plots as showed in (Table 2). All tested insecticides in (Table 2) caused significant mortality in population of cotton pink bollworm even 7th days after spray. Triazophos + Deltamethrin 40EC+2.5EC was statistically highly effective with mortality in cotton pink

bollworm population as 61.1 and 90.2% on 3rd and 7th days of treatment during 2016 followed by Gamma cyhalothrin 60CS that caused a mortality in population of cotton pink bollworm as 59.2 and 73.4. In case of Spinetoram 120SC the mortality in population as 54.3 and 71.6% after 3rd and 7th days of treatment. While in case of Deltamethrin 2.5EC and Triazophos 40EC similar effect on the mortality of cotton pink bollworm was observed even after 3rd and 7th days after treatment i.e (50.4 & 63.7) and (44.6 & 58.4) as described in table 3. The minimum control was observed by Chlorantraniliprole 20 SC as 38.8 and 53.9 on 3rd and 7th days after application. Triazophos + Deltamethrin 40EC+2.5EC proved to be the best product after 3rd and 7th days after application of insecticides. Non selective use of pesticides leads to water pollution, soil degradation, pest resistance and resurgence and ozone depletion (Naeem et al. 2012). Younis *et al.* (2007) reported that, the synthetic pyrethroid, Lambda cyhalothrin and deltamethrin exhibited the greatest reduction in bollworms infestation compared with the organophosphorus pesticide treatment.

Table 2. Mean percent population mortality of cotton Pink bollworm after application of different insecticides on cotton during 2016.

Treatments	Dose/ha (g,ml)	Av. pest population before spray	Post treatment average population/plant		Mortality (%)	
			3 th day	7 th day	3 th day	7 th day
Triazophos	2500	8	5.7e	4.7e	44.6	58.4
Deltamethrin	1000	9	5.1d	4.1d	50.4	63.7
Triazophos + Deltamethrin	2500+ 1000	7	4.0a	1.1a	61.1	90.2
Gamma Cyhalothrin	250	8	4.2b	3.0b	59.2	73.4
Spinetoram	300	9	4.7c	3.2c	54.3	71.6
Chlorantraniliprole	312	7	6.3f	5.2f	38.8	53.9
Control		8	10.3	11.3	-	-

Each value is a mean of three replications. Means sharing similar letters in columns are not significantly different byDMR test (P=0.05)

It is evident from the (Table 3) that insecticides were found to be effective in controlling population of cotton pink bollworm during 2017 under field conditions. All tested insecticides (Table 1) caused significant mortality in population of cotton pink bollworm even 7 days after spray. Triazophos + Deltamethrin 40EC+2.5EC was statistically highly effective with mortality in cotton pink bollworm population as 59.3 and 89.4% on 3rd and 7th days of treatment during 2016 followed by Gamma cyhalothrin 60CS that caused a mortality in population of cotton pink bollworm as 58.6 and 84.9%. In case of Spinetoram 120SC the mortality in population as 49.8 and 75.2% after 3rd and 7th days of treatment. While in case of Deltamethrin 2.5EC and Triazophos 40EC similar effect on the mortality of cotton pink bollworm was observed

even after 3rd and 7th days after treatment i.e (45.8 & 66.) and (45.2 & 61.2) as described in table 3. The minimum control was observed by Chlorantraniliprole 20 SC as 37.8 and 49.4% on 3rd and 7th days after application. Triazophos + Deltamethrin 40EC+2.5EC proved to be the best product after 3rd and 7th days after application of insecticides. PBW resistance to chlorinated hydrocarbons was reported in Mexico and Texas in the 1950s and 1960s (Lowry and Berger, 1965). El-Metwally et. al. (2003) showed that synthetic pyrethroids (Fenpropathrin, esfenvalerate and lambda cyhalothrin) gave the highest reduction of bollworms infestation followed by IGRs, flufenoxuron, hexaflumzuron mixed with O.P chlorpyrifos, while IGRs alone gave least reduction.

Table 3. Mean percent population mortality of cotton Pink bollworm after application of different insecticides on cotton during 2017.

Treatments	Dose/ha (g,ml)	Av. pest population before spray	Post treatment average population/plant		Mortality (%)	
			3 th day	7 th day	3 th day	7 th day
Triazophos	2500	8	4.49e	3.6e	45.2	61.2
Deltamethrin	1000	9	4.44d	3.1d	45.8	66.6
Triazophos + Deltamethrin	2500+1000	7	3.33a	0.98a	59.3	89.4
Gamma Cyhalothrin	250	8	3.39b	1.4b	58.6	84.9
Spinetoram	300	9	4.11c	2.3c	49.8	75.2
Chlorantraniliprole	312	7	5.10f	4.7f	37.8	49.4
Control		7	8.2	9.3	-	-

Each value is a mean of three replications. Means sharing similar letters in columns are not significantly different by DMR test (P=0.05)

Conclusion

It is concluded from the research trial that all the insecticides proved to be effective for controlling cotton pink bollworm but Triazophos + Deltamethrin 40EC+2.5EC @ 2500+1000 ml/ha proved to be more effective against cotton pink bollworm followed by Gamma cyhalothrin 60CS and Spinetoram 120SC.

References

1. Abd El-Mageed, A.E.M., L.R. El-Gohary and H.F. Dahi. 2007. Evaluation of several programs of sequences pesticides application on cotton bollworms and some other sucking pest in cotton field. J. Entomol., 4: 93-103.
2. Anonymous, (2015-16). "Economic survey of Pakistan", Ministry of Food and Agriculture, Islamabad. pp. 21.
3. Bariola, L. A., and T. J. Henneberry. 1980. Induction of diapause in field populations of the pink bollworm in the Western United States. Ibid. 9:376 -380.
4. Bennett, O.L., Erie, L.J. and MacKenzie, A.J. (1967) Boll @ber and spinning properties of

cotton as affected by management practices. US Dept Agricult. Tech. Bull. 1372, 109pp, Washington DC.

5. Burrows, T.M., V. Sevacherian, H. Browning, and J.Baritelle. 1982. The history and cost of the pink boll- worm in the Imperial Valley. Bulletin of the Entomologi- cal Society of America 28:286–290.
6. Common, I.F.B. 1958. A revision of the pink bollworms of cotton (*Pectinophora* Busck (*Lepidoptera: Gelechiidae*)) and related genera in Australia. Australian Journal of Zoology 6:268–306
7. Duncan, D.B. 1955. Multiple Ranges and Multiple F. Test Biomet. 11:1-2.
8. El-Sorady, A.E.M., A.A.S. El-Zanan, M.K.A. Abo-Sholoha and A.A. El-Dahan. 1998. Influence of some insecticide sequences on natural and artificial infestation with pink bollworm *Pectinophora gossypiella* (Saund..). Egypt, J. Agric. Res., 76(2): 585-596.
9. El-Metwally, H.E., S.A. El-Mahy, Alia Abdel-Haffz and R.M. Amer. 2003. Residues of esfenvalerate and flufenoxuron in cotton bolls and the relationship between pesticide dynamics and efficacy. Bull. ent. Soc. Egypt., Econ. Ser., 29: 199-210.

10. Flint HM, Yamamoto AK, Parks NJ, Nyomura K (1993) Aerial concentrations of gossyplure, the sex pheromone of the pink bollworm (Lepidoptera: Gelechiidae), within and above cotton fields treated with long-lasting dispensers. *Environmental Entomology* 22: 43-48.
11. Gahukar, R.T. 2006. Improving the conservation and effectiveness of arthropod parasitoids for cotton pest management. *Outlook on Agric.*35 (1): pp 41-49
12. Ghure, S.T., B.S. Kharbde and D.S. Patil. 2008. Bioefficacy of new pesticides against bollworm complex of cotton (*Gossypium* spp.). *International J. of Plant Protection* 1(2): 106-109.
13. Gosalwad, S.S., S.K. Kamble, D.W. Wadnerkar and K.B. Awaz. 2009. Efficacy of some newer insecticides for control of cotton bollworms. *Journal of Cotton Research and Development*, 23(2): 282-285.
14. Greenberg SM, Adamczyk JJ, Armstrong JS. 2012. principles and practices of integrated pest management on cotton in the Lower Rio Grande Valley of Texas, *Integrated Pest Management and pest Control-current and future tactics*, Dr. Sonia Soloneski (Ed.), ISBN: 978-953-51-0050-8, In Tech, Available from: www.intechopen.com
15. Haynes, K.F., Miller, T.A., Staten, R.T., Li, W.G. and Baker, T.C. (1986) Monitoring insecticide resistance with insect pheromones. *Experientia* 42, 1293±5.
16. Haynes, K.F., Miller, T.A., Staten, R.T., Li, W.G. and Baker, T.C. (1987) Pheromone trap for monitoring insecticide resistance in the pink bollworm (Lepidoptera: Gelechiidae) *Newton* for resistance management. *Environ. Entomol.* 16, 84±9.
17. Henneberry TJ, Clayton TE (1982) Pink bollworm of cotton (*Pectinophora gossypiella* (Saunders)): male moth catches in gossyplure-baited traps and relationships to oviposition, boll infestation and moth emergence. *Crop Protection* 1: 497-504.
18. Henderson, C. F. nd E. W. Tilton, 1955. Tests with acaricides agaainst the brow whet mite, *J. Econ. Entomol.* 48: 157-161.
19. Hussein N.M.;Shalaby F.F.; EL-Khayat E.F.;Tawfik S.M. and Salem M.S. (2002).Effect of certain agrochemicals on cotton a growth and bollworms infestation during three successive seasons at Kalubia Governorate, Egypt.Proceedings of the 2nd International Conference, Plant Protection Research Institute, December 21-24, 2002 Cario, Egypt, pp 854-865.
20. Hutchison WD, Beasley CA, Henneberry TJ, Martin JM (1988) Sampling pink bollworm (Lepidoptera: Gelechiidae) eggs: potential for improved timing and reduced use of insecticides. *Journal of Economic Entomology* 81: 673-678.
21. Hutchison WD, Beasley CA, Henneberry TJ, Martin JM (1991) Timing insecticide applications for pink bollworm (Lepidoptera: Gelechiidae) management: Comparison of egg and larva treatment thresholds. *Journal of Economic Entomology* 84: 470-475.
22. Khidr, A.A., W.M.H. Desuky, A.A. El-Sheakh and S.A. Raslan. 1996. Sequential use of some insecticides against cotton bollworms in control trials. *Egypt, J. Agric. Res.*, 74(2): 321-331.
23. Lowry, W.L. and Berger, R.S. (1965) Investigations of pink bollworm resistance to DDT in Mexico and the United States. *J. Econ. Entomol.* 58, 590±1.
24. Mohyuddin, A.I., G. Jilani, A.G. Khan, A.I. Humza and Z. Mehmood, (1997). "Integrated pest management of major cotton pests by conservation, redistribution and augmentation of natural enemies", *Pakistan J Zool.*, 29(3): pp 293.298.
25. Naeem, M., Farid, A. Khan, M.H. and Ali S.K. (2012). Laboratory studies on the comparative effect of neem oil (*Azadirachta indica*) and insecticides on *Trichogramma chilonis* (Ishii). *Pak. J. Entomol.* Karachi, 27(1): pp 33-38.
26. Ozyigit, I.I., M.V. Kahraman and O. Ercan. 2007. Relation between explants age, total phenols and regeneration response in tissue cultured cotton (*Gossypium hirsutum* L.). *Afric. J. Biotech.* 6(1): pp 003-008.

27. Schwartz, P. H. 1983. Losses of yield in cotton due to insects. In: Agricultural Handbook. US Department of Agricultural Research Service. Beltsville, Maryland.
28. Shaaban, A.M. and H.S. Radwan. 1974. Population dynamics of pink bollworm (*Pectinophora gossypiella*) (Saund) in relation to blooming and fruiting curves of cotton plants. *Pf, Kranth*, 4: 206-211.
29. Tamarak, S. A. 2007. Suscetibility of Spodoptera littoralis to old and new generation of Spinosyn products in five cotton. *Resistant pest mangament* ,vol 16,No.2
30. Vanilla S, Biradar VK, Sabesh M, Bambawale OM. 2007. Know your cotton insect pests: Spotted and spiny bollworms. *Crop protection folder series*: 5 of 11.
31. Wilson, A.G.L. 1972. Distribution of pink bollworm, *Pectinophora gossypiella* (Saund.), Australia and its status as a pest in the Ord irrigation area. *Journal of the Australian Institute of Agricultural Science* 38:95–99.
32. Watson, T.F., and D.G. Fullerton. 1969. Timing of insecticide applications for control of pink bollworm in Arizona. *Journal of Economic Entomology* 62:682–685.
33. Younis, A. M.; Hamouda, H. H. S.; Ibrahim, A. S. and Zeitoum, M. A. Z. (2007). Field evaluation of certain pesticides against the cotton bollworms with special reference to their negative impact on beneficial arthropoda .*African Crop Science conference Proceedings 8th African Crop Science Society, Elminia, Egypt, October 27-31*, pp. 993-1002.

Access this Article in Online	
	Website: www.darshanpublishers.com
	Subject: Agricultural Sciences
Quick Response Code	
DOI: 10.22192/ijcrbs.2018.05.07.002	

How to cite this article:

Ashiq Hussin Sanghi, Tanweer Ahmed, Muhammad Aslam, Laila Khalid and Asma Aslam. (2018). Efficacy of different insecticides against pink bollworm *Pectinophora gossypiella* (Saund.) (Lepidoptera:Gelechiidae) on cotton crop in ecological zone Of rahim yar Khan.. *Int. J. Compr. Res. Biol. Sci.* 5(7): 15-22.

DOI: <http://dx.doi.org/10.22192/ijcrbs.2018.05.07.002>